

Maternofetal Thyroid Action and Brain Development

R.G. Ahmed

Division of Anatomy and Embryology, Zoology Department, Faculty of Science, Beni-Suef University, Beni-Suef, Egypt

Email: r_g_a_ahmed@yahoo.com & ahmedragab08@gmail.com

Tel. number: 002-010-9147-1828

Editorials and Commentary

Thyroid hormones (THs) are a key regulatory factor of the developmental program and maternal-fetal communication network. The cellular basis for these effects lies in the organizational role of TH in neuronal migration, synaptogenesis and differentiation of multiple cell types. Thus, any vigorous changes in the THs levels during the development may cause a pathophysiological states and serious damage to the structural development and organization of the brain. It can be hypothesized that the disturbance in the maternofetal THs due to antithyroid drugs, Levo-thyroxine or environmental disruptors may lead, in turn, to the biochemical dysfunctions, morphofunctional alterations, developmental abnormalities and brain damage if not corrected prior/after the birth. Thus, further studies need to be done to emphasize this concept.



Council for Innovative Research

Peer Review Research Publishing System

Journal of Advances in Biology

Vol. 7, No. 1

editorsjab@gmail.com, editor@cirjab.com



I- Normal state

Due to TH action can be controlled in individual cells through selective TH uptake and intracellular TH metabolism, the placenta is an important axis in the maternal-fetal communication system for THs which are vital for the normal development and differentiation of the fetus (Ahmed et al., 2008; Scapin et al., 2010; Ahmed, 2012a; Forhead and Fowden, 2014; Micke et al., 2015). In general, intracellular activation or inactivation of L-thyroxine (T4) and 3,5,3'-triiodothyronine (T3) in turn is determined by three types of iodothyronine deiodinases (Ds), namely D1, D2, and D3 (Incerpi et al., 2005; Gereben et al., 2008; Horn and Heuer, 2010; Van Herck et al., 2012; Chung, 2014; Guzmán-Gutiérrez et al., 2014; Sánchez-Huerta et al., 2015). I reported that the placenta transports and metabolizes maternal THs. and mainly expresses D3, which inactivates T4 and other iodothyronines and thus limits the transport of maternal active THs to the fetus in the late pregnancy (Ahmed, 2012b). D2 is also dynamic in the placenta and locally provides active T3 from the maternal T4 for placental metabolic functions (Áhmed et al., 2008; Ahmed, 2012a). The placental expression of D1, which also activates T4 to T3, is still controversial. Additionally, the ability to transport THs in and out of cells has been described in members of different transmembrane TH-transporters (THTs) including the monocarboxylate transporters (MCT), L-type amino acid transporters (LAT), Na+/Taurocholate cotransporting polypeptide (NTCP) and organic anion transporting polypeptides (OATP) (Yen, 2001; Incerpi et al., 2002; Suzuki and Abe, 2008; Visser et al., 2008; Loubiére et al., 2010; Guzmán-Gutiérrez et al., 2014). To date six different THTs are recognized to be present in the placenta: MCT8, MCT10, LAT1, LAT2, OATP1A2 and OATP4A1 but their relative contributions to placental TH transport are unknown (Patel et al., 2003). There is elevation in the MCT8 mRNA expression with the gestation progress (Chan et al., 2006) but there is limited information regarding the ontogeny of the other THTs. Moreover, placental membranes are also concerned in 4'-OHsulfation reactions of iodothyronines (Köhrle, 1997). Sulfation plays a role in TH metabolism by cooperating between the deiodination and sulfation pathways of TH (Kilby et al., 2005).

The cellular action of THs is frequently classified as genomic (nuclear) and non-genomic (initiated either at cytoplasm or at membrane TH receptors) (Incerpi, 2011; Ahmed et al., 2013). Because integrin ανβ3 contains a cell surface receptor for TH but also is a co-receptor for insulin-like growth factor type 1 (IGF1) (Clemmons and Maile, 2003), we postulated that TH might modulate IGF-I actions (Incerpi et al., 2014). The proliferative actions of T4 initiated at integrin ανβ3 are MAPK dependent; T4 is anti-apoptotic and pro-angiogenic (Lin et al., 2013; Davis et al., 2014). TH also modulates the actions of interferon-γ (Lin et al., 1996), adiponectin (Wang et al., 2008, Ahmed, 2013 & 2014), leptin (Ahmed, 2013 & 2014), tumor necrosis factor-α (TNF-α) (Ahmed, 2013; Ahmed et al., 2013), brain-derived neurotrophic factor (BDNF), neurotrophin (NT)-3, NT-4/5, inositol trisphosphate oic acid receptor (ROR) a, myelin basic protein (MBP) (Yu et al., 2015), and growth f (IP) 3 receptor s, such as epidermal ransforming growth factor-a (TGF-a) (Shih et al., 2004), vascular growth factors (Date growth factor (Yu et al., 2015) by non-genomic mechanisms. An alternative clarific prectin, and TNF-a are concerned in the modulation of thyroid function and insulin substituting hormone (TSH), growth hormone (GH), and GH/IGF axis (Ahmed, 2013) and the concentrations of these markers is synergistic and intimately interconnected the standard axis (LIDTA) and CH/IGF axis desired to the concentrations. et al., 2004; Mousa growth factor), transforming growth factor-o et al., 2006) a rve n of my recent work is that the lep dipo livity, and they may interrelate wi roid an be deduced that the steady i the action of the -thyroid axis (HPTA) and GH/IGF1 axis during the development. Furthermore, bind f T3 to its nuclear hypothalami (TRs) directly influences transcription of numerous genes that are significant in dev thyroid rece nent (Harvey CB, There are growing data that T4 could affect nuclear T3 action by causing alteration of TH nuclear receptors ion (Hung-Yun et al., 2005) and phosphorylation (Davis et al., 2000; Babu et al., 2011). Notably, placental cells inity, stereo-specific, energy-dependent uptake systems for T4 and T3. Williams, 2 through ac inity express hi

II- Abnormal state

on is accompanied by deep alterations in the thyroidal economy (hypo- or hyper-thyroidism), bination of factors specific to the pregnant case, which together agree to stimulate the maternal thyr G esulting from a complex id axis (Ahmed Ahmed, 2011). The low maternal levels of T4 seen in gestation period may be compensated by et al., 20 gher placental ., 2014). High via elevation in the activity of placental TH transport and metabolism (Guzmán-Gut availabil during the first half of gestation were related to lower birth weight and ing that maternal thyroid function may influence the fetal growth, even w all gestational age newborns, suggesting n the normal range (León et al., 2015). Moreover, clinical studies demonstrated that m cit during the first trimester of pregnancy can influence the result of human neurodevelopment (Pop et al., 1999 & 2003). Experiments in rats proved that early maternal TH deficiency affects neuronal migration in the cortex (Lavado-Autric et al., 2003), while maternal hyperthyroidism too can disturb the fetal developing brain (Evans et al., 2002). In chick, we reported that the maternal transport of methimazole (MMI) and its metabolites to the embryo can be injurious for embryonic development, in particular, for the brain, by a combination of anti-thyroidal and probably local cytotoxic effects, an observation that may also be of significance to the human condition (Van Herck et al., 2013). These alterations may be either directly or indirectly linked to TH action. As well, the clinical epidemiological and basic observations obviously demonstrate that maintaining normal TH regulation from the starting of gestation is significant to diminish the risk of obstetric complications and to ensure optimal neurodevelopment of the offspring (Ahmed, 2012a). Thus, the suggestion that neurodevelopmental deviations might be associated to the THs is reasonable. More importantly, the THs disorders due to antithyroid drugs (MMI or propylthiouracil (PTU)), and polychlorinated biphenyls (PCBs) or 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) during the development may lead to deformations rather than coordinated shifts in the relative development of numerous central biological organizations that guides to a multitude of irreversible morphological and physiological irregularity (pathophysiological and patho-developmental conditions) (Figure 1) (Ahmed, 2011, 2012a, 2013 & 2014; Ahmed et al., 2014; Ahmed and El-Gareib, 2014). These can be ascribed to the diminution in the level of GH (Ahmed, 2013), the disruption of the activities of THs (Ahmed, 2012a), the synthesis and release of growth hormone-releasing hormone (GH-RH), the sensitivity of the pituitary gland to GH-RH, and the transcription of the GH gene (Osfor et al., 2013). It has also been shown that the reduction in the concentration of IGF1 was related to the variations in the activities of THs and insulin that may delay growth (Ahmed, 2013). This status might be elucidated by the depletion in leptin turnover and degradation (Houseknecht et al., 1996) or stimulation of the HPTA in response to



low TH concentrations in rats, dogs, and humans (Mazaki-Tovi et al., 2010). Especially, the loss of body weight may propose a decline in the general health level of animals (Fernandes et al., 2007), which can be vital in the explanation of thyroid effects (Ahmed et al., 2008; Ahmed, 2012a, 2013 & 2014). This is possibly due to the disorder of the hormonal homeostatic mechanisms during development. In pathological/abnormal pregnancies with either maternal or fetal THs disorders (hypo- or hyper-thyroidism), the placenta lacks the full compensatory roles essential to optimize the maternal-fetal transport of THs to achieve the normality of TH levels in the fetus (Ahmed, 2012b). This difference could be due to the experimental models, developmental period, and animal species used. Importantly, the actions of THs are highly pleiotropic, affecting many tissues at different developmental periods. Also, MCT8 insufficiency has essential metabolic consequences in the brain that could not be associated to deficiency or overload of TH supply to the brain during maturity. More recently, THTs and Ds are important regulators of intracellular T3 availability and therefore contribute to the control of TRs-dependent CNS development and early embryonic life (Ahmed, 2015). As a result, their effects on proliferation and differentiation are extremely heterogeneous depending on the cell type, the cellular context, and the developmental or transformation condition. During brain development, iodine deficit, maternal thyroid dysfunction, interruption of the maternal transfer of T4 and neonatal thyroid distortion together with the genetic factors contribute to neurodevelopmental deficits. This leads to irreversible variations and harm to the fetal CNS (i.e. abnormal corticogenesis). Interestingly, even mild to moderate maternal hypothyroxinemia may result in suboptimal neurodevelopment (Morreale de Escobar et al., 2007) and may affect the neuropsychological development of the child (Gärtner, 2009). Maternal subclinical hypothyroidism had major negative impact on neurodevelopment (Zhang et al., 2014). In addition, maternal thyroid dysfunction, hyper- and hypothyroidism, would increase the risk of seizure in the child via slight alter in brain structure (Andersen et al., 2013). Both hypothyroid and hyperthyroid diseases (resulting from genetic and acquired aetiologies) can lead to characteristic neurological diseases, with cognitive delay, extrapyramidal movement disorders, neuropsychiatric indications, and neuromuscular demonstrations (Kurian et al., 2014). As well, defects in fetal Ds or THTs may have more impact on fetal brain since they can result in intracellular T3 deficiency despite sufficient maternal TH supply (Ahmed, 2015). These deficiencies can adversely affect neurodevelopment. These results imply that the disturbance in the maternofetal THs, Ds, THTs or TRs may impair the fetal neuroendocrine system and delay the development (Figure 2).

Finally, all these abnormalities can be prevented (1) by maternal thyroxine therapy; and (2) when a normal iodine supply is ed to the mother before and during pregnancy and to the neonate and young infant during the vital period of brain opment. Inadequate iodine intake in the mother can cause low T4 in gestation and also insufficient production of T4 in fed infants when enough T4 is crucial for normal brain development. It therefore follows that mothers should be screened for provided to the development. I breast-fed infa s probable during gestation. An sufficient serum concentration of T4 is essential for hypothyroidisn treated before or as early a better understanding of these mechanisms would <mark>also allow us to refine any deviations thway of TH action, e.g., T3/rT3 ratio, deiodinase or transporter polymorphisms which p</mark> developing bra or polymorphisms in athway of TH action, e.g., elements of t edict the emotional ssociate with other potentially TH related effects. I hope that new insights into the complex reaction to Th actions by THs and their receptor trol cell proliferation and differentiation will be provided in the near future.

Author Disclosure Statement: The author has nothing to disclose.

Funding: This work was supported by a National Grant from the Research Center, Beni-Suef University, Beni-Suef city, Egypt.

References

- Ahmed OM, El-Gareib AW, El-bakry AM, Abd El-Tawab SM, Ahmed RG. Thyroid hormones states and brain development interactions. Int. J. Dev. Neurosci. 2008;26 (2):147–209. Review.
- 2. Ahmed RG, El-Gareib AW, Incerpi S. Lactating PTU exposure: II- Alters thyroid-axis and prooxidant-antioxidant balance in neonatal cerebellum.Int. Res. J. of Natural Sciences 2014;2(1):1-20.
- 3. Ahmed RG, El-Gareib AW. Lactating PTU exposure: I- Alters thyroid-neural axis in neonatal cerebellum. Eur. J. of Biol. and Medical Sci. Res. 2014;2(1):1-16.
- 4. Ahmed RG, Paul J. Davis, Faith B. Davis, Paolo De Vito, Ricardo N. Farias, Paolo Luly, Jens Z. Pedersen, Sandra Incerpi. Non-genomic actions of thyroid hormones: from basic research to clinical applications. An update. Immunology, Endocrine & Metabolic Agents in Medicinal Chemistry, 2013;13(1):46-59.
- 5. Ahmed RG. Editorial: Do PCBs modify the thyroid-adipokine axis during development? Annals Thyroid Res. 2014;1(1):11-12.
- 6. Ahmed RG. Early weaning PCB 95 exposure alters the neonatal endocrine system: thyroid adipokine dysfunction. Endocrinology, 2013;219(3):205-215.
- 7. Ahmed RG. Hypothyroidism and brain developmental players. Thyroid Res. J. 2015 (In press).
- Ahmed RG. Maternal-fetal thyroid interactions. In the Thyroid Hormone, Chapter 5. Ed N.K. Agrawal. Croatia: In tech Open Access Publisher, University Campus, STeP Ri Slavka Krautzeka 83/A 51000 Rijeka, 2012b. pp. 125-156.
- 9. Ahmed RG. Maternal–newborn thyroid dysfunction. In Developmental Neuroendocrinology. Ed RG Ahmed. LAP LAMBERT Academic Publishing GmbH & Co KG: Saarbrücken, Germany. 2012a. pp. 1–369.



- Ahmed RG. Perinatal TCDD exposure alters developmental neuroendocrine system. J. of Food and Chemical Toxicol. 2011;49:1276-1284.
- 11. Andersen SL, Laurberg P, Wu CS, Olsen J. Maternal thyroid dysfunction and risk of seizure in the child: A Danish nationwide cohort study. Journal of Pregnancy 2013; 1-10.
- 12. Babu S, Sinha RA, Mohan V, Rao G, Pal A, Pathak A, Singh M, Godbole MM. Effect of hypothyroxinemia on thyroid hormone responsiveness and action during rat postnatal neocortical development. Experimental Neurology 2011;228: 91–98.
- 13. Chan SY, Franklyn JA, Pemberton HN, Bulmer JN, Visser TJ, McCabe CJ. Monocarboxylate transporter 8 expression in the human placenta: the effects of severe intrauterine growth restriction. J Endocrinol 2006;189: 465–471.
- 14. Chung HR. Adrenal and thyroid function in the fetus and preterm infant. Korean J Pediatr 2014;57(10):425-433.
- 15. Clemmons DR, Maile LA. Integral membrane proteins that function coordinately with the insulin-like growth factor I receptor to regulate intracellular signaling. Endocrinology 2003;144:1664–1670.
- 16. Davis FB, Mousa SA, O'Connor L, Mohamed S, Lin HY, Cao HJ, Davis PJ. Proangiogenic action of thyroid hormone is fibroblast growth factor-dependent and is initiated at the cell surface. Circ Res 2004;94: 1500–1506.
- 17. Davis PJ, Hercbergs A, Luidens MK, Lin H-Y. Recurrence of differentiated thyroid carcinoma during full TSH suppression: Is the tumor now thyroid hormone dependent? J. Horm. Canc. 2014 (in press).
- 18. Davis PJ, Shih A, Lin HY, Martino LJ, Davis FB. Thyroxine promotes association of mitogen-activated protein kinase and nuclear thyroid hormone receptor (TR) and causes serine phosphorylation of TR. J. Biol. Chem. 2000;275: 38032–38039.
- 19. Evans IM, Pickard MR, Sinha AK, Leonard AJ, Sampson DC, Ekins RP. Influence of maternal hyperthyroidism in the rat on the expression of neuronal and astrocytic cytoskeletal proteins in fetal brain. Endocrinol. 2002;175:597-604.
- 20. Fernandes GS, Arena AC, Fernandez CD, Mercadante A, Barbisan LF, Kempinas WG. Reproductive effects in male rats exposed to diuron. Reproductive Toxicology 2007;23:106–112.
- 21. Forhead AJ, Fowden AL. Thyroid hormones in fetal growth and prepartum maturation. Journal of Endocrinology 2014;221: R87–R103.
- 22. Gärtner R. Thyroid diseases in pregnancy. Current Opinion in Obstetrics and Gynecology 2009;21: 501-507.
- 23. Gereben B, Zeold A, Dentice M, Salvatore D, Bianco AC. Activation and inactivation of thyroid hormone by deiodinases: local action with general consequences. Cell. Mol. Life Sci. 2008;65:570–590.
- 24. Guzmán-Gutiérrez E, Veas C, Leiva A, Escudero C, Sobrevia L. Is a low level of free thyroxine in the maternal circulation associated with altered endothelial function in gestational diabetes? Front Pharmacol. 2014 Jun 6.5:136. doi: 10.3389/fphar.2014.00136. eCollection 2014.
- 25. Harvey CB, Williams GR. Mechanism of thyroid hormone action. Thyroid 2002;12:441-446.
- 26. Horn S, Heuer H 2010. Thyroid hormone action during brain development: more questions than answers. Mol. and Cell. Endocrinol. 2010;315:19–26.
- 27. Houseknecht KL, Mantzoros CS, Kuliawat R, Hadro E, Flier JS, Kahn BB. Evidence for leptin binding to proteins in serum of rodents and humans: modulation with obesity. Diabetes 1996;45:1638–1643.
- 28. Hung-Yun L, Hopkins R, Cao HJ, rang HY, Alexander C, Davis FB, Davis PJ. Acetylation of nuclear hormone receptor superfamily members: thyroid hormone causes acetylation of its own receptor by a mitogen-activated protein kinase dependent mechanism. Steroids 2005;70: 444–449.
- 29. Incerpi S, De Vito P, Luly P, Spagnuolo S, Leoni S. Short-term effects of thyroid hormones and 3,5-diiodothyronine on membrane transport systems in chick embryo hepatocytes. Endocrinol. 2002;143:1660-1668.
- 30. Incerpi S, Hsieh M-T, Lin H-Y, Cheng G-Y, De Vito P, Fiore AM, Ahmed RG, Salvia R, Candelotti E, Leone S, Luly P, Pedersen JZ, Davis FB, Davis PJ. Thyroid hormone inhibition in L6 myoblasts of IGF-I-mediated glucose uptake and proliferation: new roles for integrin ανβ3. Am. J. Physiol. Cell Physiol. 2014;307:C150–C161.
- 31. Incerpi S, Scapin S, D'Arezzo S, Spagnuolo S, Leoni S. Short-term effect of thyroid hormone in prenatal development and cell differentiation. Steroids 2005;70:434-443.
- 32. Incerpi S. Editorial. Non-genomic effects of thyroid hormones in skeletal muscle and central nervous system: from zebrafish to man. Immun. Endocr. Metab. Agents Med. Chem. 2011;11:150-151.
- 33. Kilby MD, Barber K, Hobbs E, Franklyn JA. Thyroid hormone action in the placenta. Placenta 2005;26:105-113.
- 34. Köhrle J. Transfer and metabolism of thyroid gland hormones in the placenta. Acta Med. Austriaca 1997;24(4):138-143.



- 35. Kurian MA, Jungbluth H. Genetic disorders of thyroid metabolism and brain development. Developmental Medicine & Child Neurology 2014;56: 627–634.
- 36. Lavado-Autric R, Auso E, Garcia-Velasco JV, Arufe Mdel C, Escobar del Rey F, Berbel P, Morreale de Escobar G. Early maternal hypothyroxinemia alters histogenesis and cerebral cortex cytoarchitecture of the progeny. J. Clin. Invest. 2003;111:1073–1082.
- 37. León G, Murcia M, Rebagliato M, Álvarez-Pedrerol M, Castilla AM, Basterrechea M, Iñiguez C, Fernández-Somoano A, Blarduni E, Foradada CM, Tardón A, Vioque J. Maternal thyroid dysfunction during gestation, preterm delivery, and birthweight. The Infancia y Medio Ambiente Cohort, Spain. Paediatr Perinat Epidemiol. 2015 Jan 7. doi: 10.1111/ppe.12172. [Epub ahead of print].
- 38. Lin HY, Thacore HR, Davis FB, Davis PJ. Potentiation by thyroxine of interferon-γ-induced antiviral state requires PKA and PKC activities. Am J Physiol Cell Physiol 1996;271: C1256–C1261.
- 39. Lin YH, Liao CJ, Huang YH, Wu MH, Chi HC, Wu SM, Chen CY, Tseng YH, Tsai CY, Chung IH, Wu TI, Tsai MM, Lin CD, Lin JH. Thyroid hormone receptor represses miR-17 expression to enhance tumor metastasis in human hepatoma cells. Oncogene 2013;32: 4509–4518.
- 40. Loubiére LS, Vasilopoulou E, Bulmer JN, Taylor PM, Stieger B, Verrey F, McCabe CJ, Franklyn JA, Kilby MD, Chan S-Y. Expression of thyroid hormone transporters in the human placenta and changes associated with intrauterine growth restriction. Placenta 2010;31: 295–304.
- 41. Mazaki-Tovi M, Feuermann Y, Segev G, Klement E, Yas-Natan E, Farkas A, Kol A, Shamay A. Increased serum leptin and insulin concentrations in canine hypothyroidism. Veterinary Journal 2010;183:109–114.
- 42. Micke GC, Sullivan TM, Kennaway DJ, Hernandez-Medrano J, Perry VE. Maternal endocrine adaptation throughout pregnancy to nutrient manipulation: Consequences for sexually dimorphic programming of thyroid hormones and development of their progeny. Theriogenology 2015;83: 604–615.
- 43. Morreale de Escobar G, Obregón MJ, Escobar del Rey F. lodine deficiency and brain development in the first half of pregnancy. Public Health Nutrition 2007;10(12A): 1554–1570.
- 44. Mousa SA, Davis FB, Mohamed S, Davis PJ, Feng X. Pro-angiogenesis action of thyroid hormone and analogs in a three-dimensional in vitro microvascular endothelial sprouting model. Int Angiol 2006;25:407–413.
- 45. Osfor MMH, Elmadbouly MA, Elsoadaa SS, Metair AH, Hussain AAA. Relation between hypercholesterolemia and insulin like growth factor-1 in elderly women suffer from hypothyroidism. Journal of Natural Sciences Research 2013;3:160–164.
- 46. Fatel P, Weerasekera N, Hitchins M, Boyd CA, Johnston DG, Williamson C. Semi-quantitative expression analysis of MDR3, FIC1, BSEP, OATP-A, OATP-C, OATPD, OATP-E and NTCP gene transcripts in 1st and 3rd trimester human placenta. Placenta 2003;24: 39–44.
- 47. Pop VJ, Brouwers EP, Vader HL, Vulsma T, van Baar AL, de Vijlder JJ. Maternal hypothyroxinaemia during early pregnancy and subsequent child development: a 3-year follow-up study. Clin. Endocrinol. 2003;59:282-288.
- 48. Pop VJ, Kuijpens JL, van Baar AL, Verkerk G, van Son MM, de Vijlder JJ, Vulsma T, Wiersinga WM, Drexhage HA, Vader HL. Low maternal free thyroxine concentrations during early pregnancy are associated with impaired psychomotor development in infancy. Clin. Endocrinol. (Oxf) 1999;50:149-155.
- 49. Rodrigues TB, Ceballos A, Grijota-Martínez C, Nuñez B, Refetoff S, Cerdán S, Morte B, Bernal J. Increased oxidative metabolism and neurotransmitter cycling in the brain of mice lacking the thyroid hormone transporter Slc16a2 (Mct8). PLoS ONE 2013;8(10): e74621. doi:10.1371/journal.pone.0074621.
- 50. Sánchez-Huerta K, Pacheco-Rosado J, Gilbert ME. Adult onset-hypothyroidism: alterations in hippocampal field potentials in the dentate gyrus are largely associated with anaesthesia-induced hypothermia. J Neuroendocrinol. 2015;27(1): 8-19.
- 51. Scapin S, Leoni S, Spagnuolo S, Gnocchi D, De Vito P, Luly P, Pedersen JZ, Incerpi S. Short-term effects of thyroid hormones during development: Focus on signal transduction. Steroids 2010;75:576–584.
- 52. Shih AI, Zhang S, Cao HJ, Tang HY, Davis FB, Davis PJ, Lin HY. Disparate effects of thyroid hormone on actions of epidermal growth factor and transforming growth factor-a are mediated by 3',5'-cyclic-adenosine 5'-monophosphate-dependent protein kinase II. Endocrinology 2004;145:1708–1717.
- 53. Suzuki T, Abe T. Thyroid hormone transporters in the brain. Cerebellum 2008;75-83.
- 54. Van Herck SLJ, Geysens S, Bald E, Chwatko G, Delezie E, Dianati E, Ahmed RG, Darras VM. Maternal transfer of methimazole and effects on thyroid hormone availability in embryonic tissues. Endocrinology 2013;218:105–115.
- 55. Van Herck SLJ, Geysens S, Delbaere J, Tylzanowski P, Darras VM. Expression profile and thyroid hormone responsiveness of transporters and deiodinases in early embryonic chicken brain development. Mol. and Cell. Endocrinol. 2012;349(2):289-297.



- 56. Visser WE, Friesema EC, Jansen J, Visser TJ. Thyroid hormone transport in and out of cells. Trends Endocrinol. Metab. 2008;19:50–56.
- 57. Wang Y, Lam KS, Yau MH, Xu A. Post-translational modifications of adiponectin: mechanisms and functional implications. Biochemical Journal 2008;409: 623–633.
- 58. Yen PM. Physiological and molecular basis of thyroid hormone action. Physiol. Rev. 2001;81(3):1097-1126.
- 59. Yu L, Iwasaki T, Xu M, Lesmana R, Xiong Y, Shimokawa N, Chin WW, Koibuchi N. Aberrant cerebellar development of transgenic mice expressing dominant-negative thyroid hormone receptor in cerebellar Purkinje cells. J. Endocrinology 2015; 1-12.
- 60. Zhang Y, Fan Y, Yu X, Wang X, Bao S, Li J, Fan C, Shan Z, Teng W. Maternal subclinical hypothyroidism impairs neurodevelopment in rat offspring by inhibiting the CREB signaling pathway. Mol Neurobiol. 2014 Sep 6. [Epub ahead of print].

Figure legends

Figure 1: Thyroid disruptors affecting the maternal-fetal thyroid alterations.

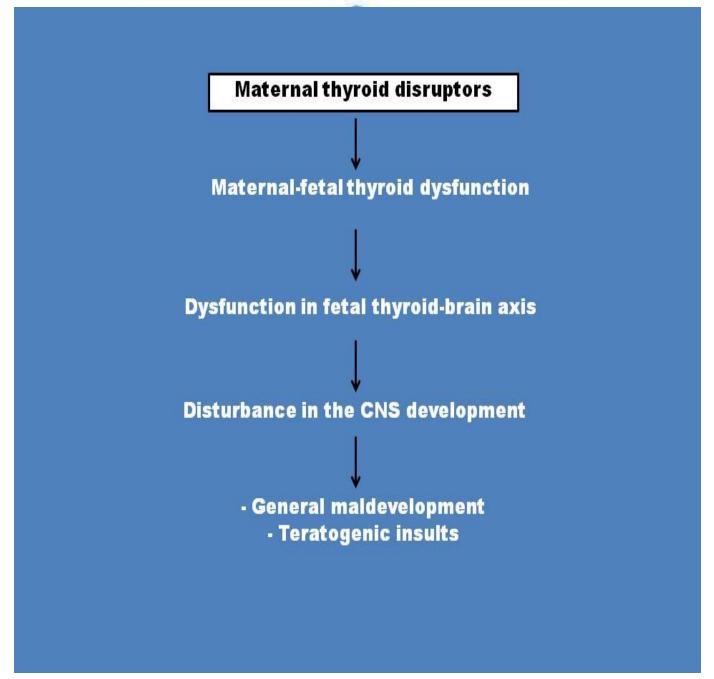




Figure 2: Schematic diagram of the maternofetal thyroid axis and the developmental neuroendocrine homeostasis.

